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Cytochrome P450 2C9 Type II Binding Studies on Quinoline-4-Carboxamide Analogues

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CYP2C9 is a significant P450 protein responsible for drug metabolism. With the increased use of heterocyclic compounds in drug design, a rapid and efficient predrug screening of these potential type II binding compounds is essential to avoid adverse drug reactions. To understand binding modes, we use quinoline-4-carboxamide analogues to study the factors that determine the structure–activity relationships. The results of this study suggest that the more accessible pyridine with the nitrogen para to the linkage can coordinate directly with the ferric heme iron, but this is not seen for the meta or ortho isomers. The π –cation interaction of the naphthalene moiety and Arg 108 residue may also assist in stabilizing substrate binding within the active-site cavity. The type II substrate binding affinity is determined by the combination of steric, electrostatic, and hydrophobicity factors; meanwhile, it is enhanced by the strength of lone pair electrons coordination with the heme iron.

Introduction

Cytochrome P450 enzymes (P450s^a) constitute a superfamily of heme-containing monooxygenases that are responsible for a large range of metabolic reactions on endogenous and exogenous substrates. Three of the major P450 isoforms involved in drug metabolism are CYP2C9, CYP2D6, and CYP3A4, which typically metabolize anions, cations, and large lipophilic molecules, respectively. Given the structural diversity of drugs, broad substrate selectivity is required for these isoforms to mediate metabolism. This makes the metabolic P450 enzymes different than other enzymes, including P450 enzymes that catalyze endogenous reactions, in that multiple metabolites are often formed from very different binding orientations.^{1,2} Even very tight binding inhibitors have been shown to bind with multiple orientations in the catalytic sites of metabolic P450 enzymes.³ Given this broad substrate selectivity, it is not surprising that potentially severe drug–drug interactions can occur due to P450s binding multiple compounds with high affinity.^{4–6} Additionally, given that the physiological concentrations of administered drugs are normally near 1 μ M, it is generally accepted that compounds that bind with a K_i less than 10 μ M may cause drug–drug interactions.

To minimize these interactions, it is important to understand the mechanisms involved in substrate binding to P450s, which are described by two binding modes called type I and type II.⁷ Type I and type II substrate binding modes have been extensively studied using UV spectroscopy,^{8–12} which characterizes spectral changes in the heme–Soret absorption band. In type I binding, the substrate binds in a hydrophobic pocket located near the catalytic site of P450. This binding displaces a water molecule that was coordinated to the catalytic face of the heme iron. The resulting five-coordinated heme iron shifts from a low-spin to high-spin state with an accompanying blue shift

in the Soret band of the absolute UV spectrum. Type I binding makes the heme more easily reduced by P450 reductase, starting the catalytic cycle of oxidative metabolism. A type II binding mode describes a situation where a substrate binds as a sixth ligand to the heme iron. In addition to the displacement of the coordinated water, the compound itself serves as a ligand. The heme iron remains in the low-spin state, which makes reduction by P450 reductase more difficult. Type II substrates typically contain an sp^2 hybridized nitrogen atom as part of a heteroaromatic system where the nitrogen lone pair coordinates with the heme iron. Type II substrates show an increase in binding affinity and results in an overall rate decrease in the catalytic cycle of oxidative metabolism. Even though binding modes are divided into two categories, the existence of mixed or multiple binding modes is inevitable. There could be an equilibrium between type I and type II binding.¹⁰

In addition to UV spectroscopy, electron paramagnetic resonance (EPR) spectroscopy has been used to differentiate between type I and type II binding modes by directly probing spin state changes in the heme iron for both bacterial and mammalian P450 enzymes.^{8,12–14} This technique provides information regarding the spin state of ferric iron, which has as many as five unpaired electrons. With dependence on substrate binding, different spin state heme signals will appear on the spectra as a result of ligand binding. A high-spin state ferric iron ($S = 5/2$) gives EPR signals as $g = 7.95, 3.78$, and 1.74 ; a low-spin state ferric iron ($S = 1/2$) gives signals as $g = 2.42, 2.25$, and 1.92 .

In order to improve metabolic stability and inhibitory potency, current practices in drug design incorporate nitrogen heterocycles (pyridines, imidazoles, and triazoles) moieties into drug molecules.^{11,15} Nitrogen incorporation has the potential of creating a type II binder that, in addition to affecting metabolic rate, could potentially inhibit coadministered drugs and lead to toxicity. Therefore, to facilitate rational drug design, it is necessary to understand and be able to predict the binding mode of a given compound as well as to balance the inhibitory potency with the metabolic stability.

Finally, some P450 enzymes are therapeutic targets. Examples include aromatase for breast cancer,¹⁶ CYP2A6 for smoking cessation,^{17,18} CYP51 for high cholesterol,^{19,20} yeast infections,

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^a Abbreviations: P450s, cytochrome P450 enzymes; EPR, electron paramagnetic resonance; 2C9, CYP2C9; 4-PI, 4-phenylimidazole; FLU, fluconazole, PDB, Protein Data Bank.

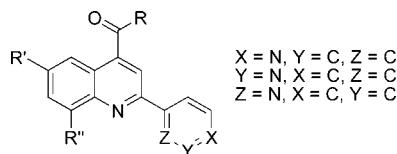


Figure 1. Quinoline-4-carboxamide analogues.

and fungal infections (*tinea pedis*, *tinea cruris*, and *candida* in HIV-AIDS).²¹ CYP51 is also a target for parasitic diseases including *American trypanosomiasis* (Chagas disease) and *African trypanosomiasis* (sleeping sickness). Many of the currently available therapeutic agents used to treat these diseases are high affinity type II binders.²¹ While type II binding enhances affinities for P450 enzymes relative to other non-P450 targets, the nonspecific inhibition of P450 enzymes using these therapeutics leads to toxicity. For example, azole antifungal agents that target CYP51 can cause birth defects similar to those observed with a phenotypic variation in the P450 reductase function that is related to a decrease in steroid metabolism.²² In order to design a viable therapeutic, the compound must specifically target P450 disease related isoforms while not inhibiting drug metabolizing P450 isoforms. Since type II binding potentially enhances binding to all P450 isoforms, it is particularly important to understand and be able to attenuate this binding interaction.

CYP2C9 (2C9) is one of the most significant drug metabolizing P450 isoforms, metabolizing many nonsteroidal anti-inflammatory drugs including ibuprofen, flurbiprofen, diclofenac, the antidiabetic sulfonylureas, and the anticoagulant (*S*)-warfarin.^{4,23,24} During routine screening of diverse inhibitors of 2C9 we found a quinoline-4-carboxamide compound that bound with a low K_i value. In an effort to elucidate the role that nitrogen coordination to the heme plays in type II substrate binding interactions with 2C9, we have synthesized a series of quinoline-4-carboxamide analogues (Figure 1 and Table 1). In addition to other structural changes, we have intentionally varied the position of the sp^2 nitrogen atom in the ring system to study heme coordination in type II binding interactions. Binding affinity, spectroscopy, computational models, and docking experiments to the known crystal structure of 2C9 are used to determine the strength and nature of these interactions. Our analogues were designed to probe binding interactions as a result of both apoprotein interactions and type II coordination while maintaining the core binding motif in order to decrease differences in binding not related to coordination to the heme iron.

Experimental Methods

Enzymes, Chemicals, and Instruments. 2C9 baculosomes were prepared as described previously.²⁵ Diclofenac and β -NADPH were purchased from Sigma-Aldrich (St. Louis, MO). HPLC-grade solvents were purchased from EMD Chemicals Inc. (Gibbstown, NJ) and used without purification. All other chemicals were purchased from commercial sources and used without purification. The 1H and ^{13}C NMR spectra were gathered on a Varian Mercury Vx 300 MHz spectrometer using TMS as an internal standard except the ^{13}C NMR spectra of compounds **19**, **20**, **21**, and **22**, which were collected by a Varian Inova 500 MHz. A Hewlett-Packard 8453 single-beam spectrophotometer with an HP 845x UV-visible system (Hewlett-Packard, Palo Alto, CA) was used to determine substrate concentrations. The HPLC system was from Agilent 1100 series with an Alltima C₁₈ 5 μ m, 3.2 mm \times 150 mm column from Alltech. Values of pK_a were estimated using MarvinSketch (ChemAxon Kft. Budapest, Hungary).

Baculosomes Incubation Conditions. Diclofenac was used as a 2C9 substrate, and K_i values were determined from the formation

rates of 4-hydroxydiclofenac. Four different final concentrations of diclofenac (5, 10, 20, and 30 μ M) in 100 mM potassium phosphate buffer at pH 7.4 were used. Three different inhibitor concentrations were added to each different concentration of diclofenac. 2C9 baculosomes (1 pmol) were added to the incubation mixtures containing the substrate and selected inhibitor and followed by preincubation for 5 min at 37 °C. NADPH was added to a final concentration of 1 mM to initiate the reaction. The final volume of all incubations was 500 μ L. Reactions were quenched after 5 min by adding 200 μ L of 6% acetic acid in acetonitrile containing an internal standard (0.05 mM phenacetin). The enzyme was removed by centrifugation, and product formation was analyzed using HPLC separation with UV detection. The HPLC system used reversed-phase chromatography beginning with 30% of mobile phase A (MeOH) and 70% of mobile phase B (30% acetonitrile/70% water containing 1 mM perchloric acid) with a linear gradient to 95% of MeOH over 6 min.²⁶ Each compound's K_i was determined from eq 1 using GraphPad Prism4 (San Diego, CA) graphing software. The low enzyme concentration of 2 nM means that substrate depletion does not need to be corrected for even for the tightest binding inhibitors.

$$v = \frac{V_m[S]}{1 + \frac{[I]}{K_m} + [S]} \quad (1)$$

K_i Determination from IC_{50} Measurements. Measurements were performed using diclofenac at K_m concentrations. Inhibitors at three different concentrations were added across to the reaction mixtures containing the same substrate concentration and P450 enzyme (1 pmol). The same incubation procedure and HPLC analysis (detailed above) were used to measure product formation rates. IC_{50} and K_i values were calculated by GraphPad Prism4 graphing software using eq 2:

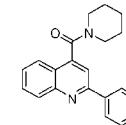
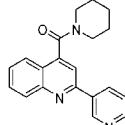
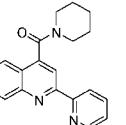
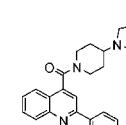
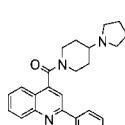
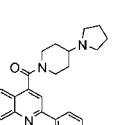
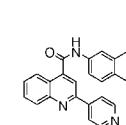
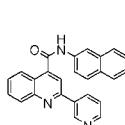
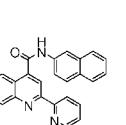
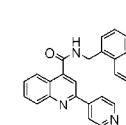
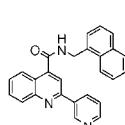
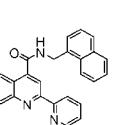
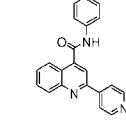
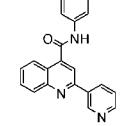
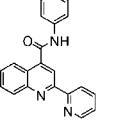
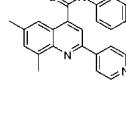
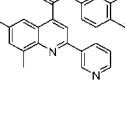
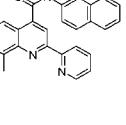
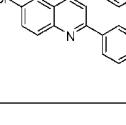
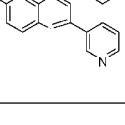
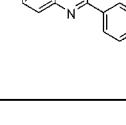
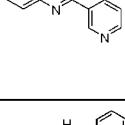
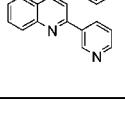
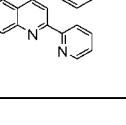
$$K_i = \frac{IC_{50}}{1 + \frac{[S]}{K_m}} \quad (2)$$

Reconstituted 2C9 System. The 2C9 protein premix was prepared as described previously²⁷ using purified 2C9 protein.²⁸ The incubation conditions were the same as the baculosomes incubation assay except 20 pmol of purified 2C9 protein was used in each sample with an incubation time of 10 min.

Synthesis of Proposed Type II Binders Based on Quinoline-4-carboxamide. All compounds were synthesized as outlined in Scheme 1 with slight modifications for each analogue.

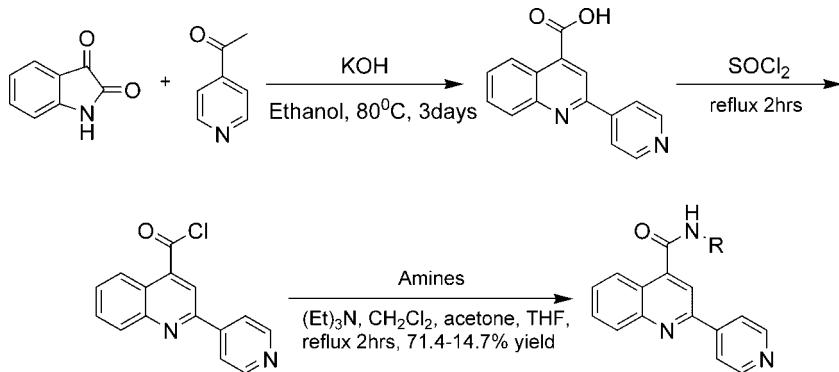
The general procedure for quinoline-4-carboxamide analogues is given with compound **7**: To a 100 mL round-bottom flask equipped with a water condenser and stir bar were combined potassium hydroxide (1.71 g, 0.030 mol) and ethanol (8 mL). The reaction mixture was stirred at 80 °C until the potassium hydroxide was completely dissolved. At this time, isatin (1.61 g, 0.011 mol) was added followed by a dropwise addition of 4-acetylpyridine (1.57 g, 0.013 mol). The mixture was refluxed with stirring at 80 °C for 3 days. At the end of this time, the reaction solvent was evaporated using a rotary evaporator, and the residue was dissolved in 50 mL of water and neutralized with 1 N HCl to pH ~6. The resulting solid was collected by vacuum filtration and washed with ethanol (25 mL \times 2), acetone (25 mL \times 2), and pentane (25 mL \times 2) to give crude 2-(pyridin-4-yl)quinoline-4-carboxylic acid, which was used directly to generate the acyl chloride in the next step. The acyl chlorides were formed by combining the crude 2-(pyridin-4-yl)quinoline-4-carboxylic acid (formed above, 500 mg, 2.00 mmol) with neat thionyl chloride (10 mL) in a 100 mL round-bottom flask fitted with an air condenser under reflux for 2 h. At the end of this time, the condenser was removed and the excess thionyl chloride was evaporated using a stream of argon to give 2-(pyridin-4-yl)quinoline-4-carboxylic chloride. The crude 2-(pyridin-4-yl)quinoline-4-carboxylic chloride (200 mg, 0.744 mmol) was dissolved in a 1:1:1 mixture of acetone/dichloromethane/triethylamine (15 mL)

Table 1. K_i Values of Type II Binding Analogues

Group	Compd	Structure (<i>para</i>)	K_i +/- SD (μ M)	Compd	Structure (<i>meta</i>)	K_i +/- SD (μ M)	Compd	Structure (<i>ortho</i>)	K_i +/- SD (μ M)
1	1		9.94 +/- 0.053	2		9.74 +/- 0.031	3		23.61 +/- 6.08
2	4		3.67 +/- 0.082	5		24.08 +/- 1.03	6		76.19 +/- 11.78
3	7		0.033 +/- 0.002	8		1.15 +/- 0.002	9		10.67 +/- 1.32
4	10		0.125 +/- 0.014	11		0.152 +/- 0.004	12		>43.5
5	13		0.10 +/- 0.008	14		4.06 +/- 1.53	15		3.47 +/- 1.40
6	16		0.011 +/- 0.001	17		8.55 +/- 1.02	18		>20.6
7	19		0.012 +/- 0.0007	20		1.89 +/- 0.20		N/A ^a	
8	21		0.009 +/- 0.001	22		>37.85		N/A ^a	
9		N/A ^a		23		1.25 +/- 0.43	24		1.32 +/- 0.10

^a Compounds not available.

Scheme 1



in a 100 mL round-bottom flask. To this flask, 2-naphthylamine (117.2 mg, 0.818 mmol) was added and the reaction mixture refluxed for 2 h. Following solvent evaporation, the crude compound was purified by chromatography (30 g, silica gel 60, 0.063–0.200 mm) using ethyl acetate. Crystallization from diisopropyl ether gave pure *N*-(naphthalen-2-yl)-2-(pyridin-4-yl)quinoline-4-carboxamide in 14.67% yield overall.

Piperidin-1-yl(2-(pyridin-4-yl)quinolin-4-yl)methanone (1). The reaction conditions were the same as above except piperidine was used instead of 2-naphthylamine to form the amide to give an overall yield of 69.5%; mp 136–137 °C; ¹H NMR (CDCl₃) δ 8.79 (dd, *J* = 1.5, 4.8 Hz, 2H), 8.23 (d, *J* = 8.4 Hz, 1H), 8.06 (dd, *J* = 1.5, 4.5 Hz, 2H), 7.87–7.78 (m, 3H), 7.63 (m, 1H), 3.90 (m, 2H), 3.17 (t, *J* = 5.7 Hz, 2H), 1.83–1.68 (m, 4H), 1.49–1.43 (m, 2H); ¹³C NMR (CDCl₃) δ 166.78, 154.51, 150.84, 148.64, 146.27, 144.61, 130.95, 130.77, 128.37, 124.86, 124.12, 121.75, 115.38, 48.54, 43.00, 26.91, 25.97, 24.61; ESI-MS *m/z* 318.5 (M + 1).

Piperidin-1-yl(2-(pyridin-3-yl)quinolin-4-yl)methanone (2). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and piperidine was used instead of 2-naphthylamine to form the amide. The overall yield was 71.4%; mp 146–147 °C; ¹H NMR (CDCl₃) δ 9.36 (dd, *J* = 0.9, 2.1 Hz, 1H), 8.72 (dd, *J* = 1.8, 4.8 Hz, 1H), 8.50 (m, 1H), 8.21 (d, *J* = 8.1 Hz, 1H), 7.86–7.76 (m, 3H), 7.60 (m, 1H), 7.47 (m, 1H), 3.90 (m, 2H), 3.18 (t, *J* = 5.4 Hz, 2H), 1.82–1.68 (m, 4H), 1.49–1.44 (m, 2H); ¹³C NMR (CDCl₃) δ 166.86, 154.67, 150.79, 149.03, 148.73, 144.48, 135.13, 134.81, 130.84, 130.54, 127.93, 124.88, 123.96, 123.67, 115.37, 48.54, 42.99, 26.91, 25.98, 24.63; ESI-MS *m/z* 318.5 (M + 1).

Piperidin-1-yl(2-(pyridin-2-yl)quinolin-4-yl)methanone (3). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine and piperidine was used instead of 2-naphthylamine to form the amide. The overall yield was 33.9%, mp 157–158 °C; ¹H NMR (CDCl₃) δ 8.73 (m, 1H), 8.66 (m, 1H), 8.51 (s, 1H), 8.21 (d, *J* = 8.7 Hz, 1H), 7.91–7.85 (m, 2H), 7.76 (m, 1H), 7.58 (m, 1H), 7.37 (m, 1H), 3.89 (m, 2H), 3.21 (t, *J* = 5.7 Hz, 2H), 1.82–1.66 (m, 4H), 1.56–1.39 (m, 2H); ¹³C NMR (CDCl₃) δ 167.20, 156.10, 155.93, 149.44, 148.32, 143.88, 137.22, 130.54, 130.33, 127.84, 125.02, 124.62, 124.54, 121.99, 115.91, 48.56, 42.90, 26.90, 26.00, 24.70; ESI-MS *m/z* 318.3 (M + 1), 233.1 (M – 85), 207.2 (M – 111).

(2-(Pyridin-4-yl)quinolin-4-yl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (4). The reaction conditions were the same as above except 4-(1-pyrrolidinyl)piperidine was used instead of 2-naphthylamine to form the amide. The crude compound was purified with chromatography using 10% MeOH in CH₂Cl₂. The overall yield was 39.4%; mp 164–165 °C; ¹H NMR (CDCl₃) δ 8.79 (s, 2H), 8.23 (d, *J* = 8.7 Hz, 1H), 8.06 (m, 2H), 7.91–7.78 (m, 3H), 7.67–7.59 (m, 1H), 4.85 (m, 1H), 3.42 (d, *J* = 13.8 Hz, 1H), 3.16–2.99 (m, 2H), 2.69 (m, 4H), 2.45–2.38 (m, 1H), 2.19–2.13 (m, 1H), 1.86–1.76 (m, 6H), 1.60–1.49 (m, 1H); ¹³C NMR (CDCl₃) δ 166.72, 154.41, 150.80, 148.50, 146.09, 144.11,

130.96, 130.65, 128.80, 124.83, 123.97, 121.70, 115.37, 61.84, 51.75, 45.93, 40.46, 31.79, 30.52, 23.33; ESI-MS *m/z* 387.5 (M + 1).

(2-(Pyridin-3-yl)quinolin-4-yl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (5). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and 4-(1-pyrrolidinyl)piperidine was used instead of 2-naphthylamine to form the amide. The crude compound was purified by chromatography using 10% MeOH in CH₂Cl₂. The overall yield was 50.2%; mp 164–165 °C; ¹H NMR (CDCl₃) δ 9.37 (dd, *J* = 1.8, 18.6 Hz, 1H), 8.73–8.71 (m, 1H), 8.53–8.47 (m, 1H), 8.23–8.19 (m, 1H), 7.92–7.77 (m, 3H), 7.67–7.56 (m, 1H), 7.49–7.45 (m, 1H), 4.90 (m, 1H), 3.46 (d, *J* = 14.4 Hz, 1H), 3.14–2.98 (m, 2H), 2.75 (m, 4H), 2.17 (m, 1H), 2.03–1.49 (m, 7H); ¹³C NMR (CDCl₃) δ 166.72, 154.59, 150.73, 148.99, 148.90, 144.17, 135.04, 134.99, 130.89, 130.48, 128.09, 124.78, 123.91, 123.57, 115.37, 61.78, 51.84, 46.06, 40.54, 32.56, 31.63, 23.37; ESI-MS *m/z* 387.4 (M + 1).

(2-(Pyridin-2-yl)quinolin-4-yl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (6). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine and 4-(1-pyrrolidinyl)piperidine was used instead of 2-naphthylamine to form the amide. The crude compound was purified by chromatography using 10% MeOH in CH₂Cl₂. The overall yield was 60.3%; mp 81–82 °C; ¹H NMR (CDCl₃) δ 8.73 (m, 1H), 8.65 (m, 1H), 8.51 (dd, *J* = 18.9 Hz, 1H), 8.21 (dd, *J* = 8.4 Hz, 1H), 7.92–7.75 (m, 3H), 7.65–7.58 (m, 1H), 7.40–7.35 (m, 1H), 4.92 (m, 1H), 3.49 (d, *J* = 12.9 Hz, 1H), 3.07–2.93 (m, 2H), 2.79 (m, 4H), 2.62–2.53 (m, 1H), 2.20–2.13 (m, 1H), 1.88–1.77 (m, 6H), 1.68–1.55 (m, 1H); ¹³C NMR (CDCl₃) δ 167.22, 156.22, 155.84, 149.51, 148.26, 143.59, 137.24, 130.49, 130.32, 128.07, 127.85, 124.96, 124.56, 121.97, 115.96, 61.98, 51.86, 46.18, 40.53, 32.67, 31.67, 23.44; ESI-MS *m/z* 387.4 (M + 1).

***N*-(Naphthalen-2-yl)-2-(pyridin-4-yl)quinoline-4-carboxamide (7).** The reaction conditions were described as above. mp 251–252 °C; ¹H NMR (DMSO) δ 11.07 (s, 1H), 8.79 (dd, *J* = 1.8, 4.8 Hz, 2H), 8.56–8.54 (m, 2H), 8.33 (dd, *J* = 1.5, 4.5 Hz, 2H), 8.24 (d, *J* = 8.4 Hz, 2H), 7.96–7.88 (m, 4H), 7.78–7.71 (m, 2H), 7.54–7.43 (m, 2H); ¹³C NMR (DMSO) δ 165.98, 154.22, 151.25, 148.58, 145.67, 144.12, 137.04, 134.01, 131.45, 130.91, 130.61, 129.22, 129.08, 128.24, 127.32, 125.89, 125.78, 124.60, 122.06, 121.08, 117.66, 117.09; ESI-MS *m/z* 376.3 (M + 1), 348.4 (M – 28), 297.4 (M – 79), 233.1 (M – 143), 205.3 (M – 171).

***N*-(Naphthalen-2-yl)-2-(pyridin-3-yl)quinoline-4-carboxamide (8).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine. The overall yield was 19.8%; mp 215–216 °C; ¹H NMR (CDCl₃) δ 9.19 (d, *J* = 1.8 Hz, 1H), 9.06 (s, 1H), 8.52–8.50 (m, 2H), 8.37 (m, 1H), 8.28 (dd, *J* = 0.6, 8.1 Hz, 1H), 8.16 (d, *J* = 8.4, 1H), 7.97 (s, 1H), 7.91–7.76 (m, 4H), 7.72 (dd, *J* = 2.1, 8.7 Hz, 1H), 7.62 (m, 1H), 7.50 (m, 2H), 7.33–7.29 (m, 1H); ¹³C NMR (CDCl₃) δ 165.81, 154.05, 150.37, 148.98, 148.55, 143.53, 135.46, 135.04, 134.41, 134.05, 131.27, 130.94, 130.40, 129.37, 128.34, 128.11, 127.92,

127.04, 125.75, 125.26, 123.95, 123.71, 120.00, 117.52, 116.29; ESI-MS *m/z* 376.5 (M + 1).

***N*-(Naphthalen-2-yl)-2-(pyridin-2-yl)quinoline-4-carboxamide (9).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (2:1). The overall yield was 23.9%; mp 261–262 °C; ¹H NMR (CDCl₃) δ 8.80 (s, 1H), 8.74–8.68 (m, 2H), 8.48–8.42 (m, 2H), 8.26–8.20 (m, 2H), 7.95–7.78 (m, 5H), 7.66–7.61 (m, 2H), 7.55–7.39 (m, 3H); ¹³C NMR (CDCl₃) δ 165.65, 155.64, 155.47, 149.16, 148.62, 142.34, 137.25, 135.03, 133.81, 131.03, 130.36, 130.25, 129.06, 128.13, 127.88, 127.66, 126.73, 125.44, 124.57, 124.46, 121.87, 119.89, 117.31, 116.18; ESI-MS *m/z* 376.5 (M + 1).

***N*-(Naphthalen-1-ylmethyl)-2-(pyridin-4-yl)quinoline-4-carboxamide (10).** The reaction conditions were the same as above except 1-naphthylmethylamine was used instead of 2-naphthylamine to form the amide. The overall yield was 72.3%; mp 188–189 °C; ¹H NMR (CDCl₃) δ 8.71 (d, *J* = 6 Hz, 2H), 8.25–8.15 (m, 3H), 7.96–7.86 (m, 5H), 7.77 (m, 1H), 7.67–7.54 (m, 4H), 7.50–7.45 (m, 1H), 6.42 (m, 1H), 5.24 (d, *J* = 5.4 Hz, 2H); ¹³C NMR (CDCl₃) δ 167.06, 154.15, 150.76, 148.93, 145.93, 143.35, 134.26, 132.88, 131.60, 130.89, 130.64, 129.45, 129.29, 128.57, 127.61, 127.18, 126.53, 125.71, 125.24, 124.23, 123.58, 121.62, 116.23, 42.81. ESI-MS *m/z* 390.5 (M + 1).

***N*-(Naphthalen-1-ylmethyl)-2-(pyridin-3-yl)quinoline-4-carboxamide (11).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and 1-naphthylmethylamine was used instead of 2-naphthylamine to form the amide. The overall yield was 10.3%; mp 175–176 °C; ¹H NMR (CDCl₃) δ 9.15 (s, 1H), 8.59 (d, *J* = 3.3 Hz, 1H), 8.38 (m, 1H), 8.21 (d, *J* = 8.1 Hz, 2H), 8.11 (d, *J* = 8.7 Hz, 1H), 7.93–7.82 (m, 3H), 7.74 (m, 1H), 7.66–7.53 (m, 4H), 7.49–7.44 (m, 1H), 7.39–7.35 (m, 1H), 6.61 (m, 1H), 5.23 (d, *J* = 5.1 Hz, 2H); ¹³C NMR (CDCl₃) δ 167.17, 154.04, 150.53, 148.86, 148.53, 143.27, 134.96, 134.34, 134.23, 133.04, 131.63, 130.72, 130.24, 129.35, 129.25, 128.06, 127.56, 127.11, 126.50, 125.70, 125.29, 123.86, 123.76, 123.66, 116.16, 42.76; ESI-MS *m/z* 390.3 (M + 1), 262.1 (M – 128), 233.0 (M – 157), 207.2 (M – 183), 141.1 (M – 249).

***N*-(Naphthalen-1-ylmethyl)-2-(pyridin-2-yl)quinoline-4-carboxamide (12).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine and 1-naphthylmethylamine was used instead of 2-naphthylamine to form the amide. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (2:1). The overall yield was 23%; mp 198–199 °C; ¹H NMR (CDCl₃) δ 8.66–8.57 (m, 3H), 8.37 (d, *J* = 7.8 Hz, 1H), 8.24–8.14 (m, 2H), 7.91–7.82 (m, 3H), 7.75 (m, 1H), 7.64–7.44 (m, 5H), 7.37–7.32 (m, 1H), 6.54 (m, 1H), 5.22 (d, *J* = 5.4 Hz, 2H); ¹³C NMR (CDCl₃) δ 167.40, 155.76, 155.69, 149.29, 148.65, 142.55, 137.33, 134.18, 133.07, 131.65, 130.36, 129.24, 129.13, 128.12, 127.41, 127.09, 126.40, 125.67, 124.79, 124.63, 123.73, 122.01, 116.35, 42.60; ESI-MS *m/z* 390.5 (M + 1).

***N*-Phenyl-2-(pyridin-4-yl)quinoline-4-carboxamide (13).** The reaction conditions were the same as above except aniline was used instead of 2-naphthylamine to form the amide. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (2:1). The overall yield was 67.8%; mp 202–203 °C; ¹H NMR (CDCl₃) δ 8.86 (s, 1H), 8.57 (d, *J* = 4.2 Hz, 2H), 8.24–8.16 (m, 2H), 7.88 (d, *J* = 6 Hz, 2H), 7.81–7.76 (m, 4H), 7.60 (m, 1H), 7.44 (t, *J* = 8.1 Hz, 2H), 7.24 (t, *J* = 7.8 Hz, 1H); ¹³C NMR (CDCl₃) δ 165.55, 153.70, 150.48, 148.89, 145.93, 143.33, 138.01, 131.02, 130.61, 129.56, 128.75, 125.50, 125.25, 124.12, 121.64, 120.39, 116.25; ESI-MS *m/z* 326.5 (M + 1).

***N*-Phenyl-2-(pyridin-3-yl)quinoline-4-carboxamide (14).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and aniline was used instead of 2-naphthylamine to form the amide. The overall yield was 41.3%; mp 215–216 °C; ¹H NMR (CDCl₃) δ 9.16 (d, *J* = 1.8 Hz, 1H), 8.82 (s, 1H), 8.52 (dd, *J* = 1.5, 4.5 Hz, 1H), 8.36 (m, 1H), 8.23 (d, *J* = 7.5 Hz, 1H), 8.15 (d, *J* = 8.4 Hz, 1H), 7.92 (s, 1H), 7.81–7.75 (m, 3H), 7.60 (m, 1H), 7.44 (t, *J* = 8.1 Hz, 2H), 7.34–7.30 (m,

1H), 7.24 (t, *J* = 7.8 Hz, 1H); ¹³C NMR (CDCl₃) δ 165.64, 154.04, 150.37, 148.95, 148.53, 143.56, 138.01, 135.02, 134.39, 130.91, 130.37, 129.55, 128.29, 125.45, 125.23, 123.94, 123.67, 120.40, 116.25; ESI-MS *m/z* 326.5 (M + 1).

***N*-Phenyl-2-(pyridin-2-yl)quinoline-4-carboxamide (15).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine and aniline was used instead of 2-naphthylamine to form the amide. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (2:1). The overall yield was 32.4%; mp 207–208 °C; ¹H NMR (CDCl₃) δ 8.75–8.66 (m, 3H), 8.38 (d, *J* = 7.8 Hz, 1H), 8.23 (d, *J* = 8.4 Hz, 1H), 8.02 (s, 1H), 7.91 (m, 1H), 7.82–7.73 (m, 3H), 7.62 (m, 1H), 7.46–7.38 (m, 3H), 7.22 (t, *J* = 7.5 Hz, 1H); ¹³C NMR (CDCl₃) δ 165.74, 155.83, 155.67, 149.37, 148.79, 142.62, 137.81, 137.46, 130.56, 130.44, 129.48, 128.31, 125.62, 125.37, 124.78, 124.65, 122.08, 120.49, 116.37; ESI-MS *m/z* 326.3 (M + 1).

6,8-Dimethyl-*N*-(naphthalen-2-yl)-2-(pyridin-4-yl)quinoline-4-carboxamide (16). The reaction conditions were the same as above except isatin was substituted with 5,7-dimethylisatin. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (1:2). The overall yield was 35.6%; mp 233–234 °C; ¹H NMR (CDCl₃) δ 8.74 (s, 1H), 8.61 (dd, *J* = 1.5, 4.8 Hz, 2H), 8.51 (d, *J* = 2.1 Hz, 1H), 7.98 (dd, *J* = 1.8, 4.8 Hz, 2H), 7.90–7.84 (m, 5H), 7.70 (dd, *J* = 2.1, 9 Hz, 1H), 7.55–7.45 (m, 3H), 2.84 (s, 3H), 2.48 (s, 3H); ¹³C NMR (CDCl₃) δ 166.35, 150.96, 150.46, 146.60, 146.27, 142.61, 138.88, 138.29, 135.46, 134.05, 133.42, 131.25, 129.35, 128.09, 127.91, 127.04, 125.74, 124.20, 121.72, 121.42, 119.99, 117.46, 115.59, 22.24, 18.31; ESI-MS *m/z* 404.5 (M + 1).

6,8-Dimethyl-*N*-(naphthalen-2-yl)-2-(pyridin-3-yl)quinoline-4-carboxamide (17). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and 5,7-dimethylisatin was used instead of isatin. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (1:2). The overall yield was 13.9%; mp 224–225 °C; ¹H NMR (CDCl₃) δ 9.23 (d, *J* = 1.8 Hz, 1H), 9.11 (s, 1H), 8.52–8.46 (m, 2H), 8.35 (m, 1H), 7.90–7.83 (m, 5H), 7.72 (dd, *J* = 2.1, 8.7 Hz, 1H), 7.54–7.43 (m, 3H), 7.29–7.25 (m, 1H), 2.75 (s, 3H), 2.46 (s, 3H); ¹³C NMR (CDCl₃) δ 166.53, 151.11, 149.90, 148.45, 146.54, 142.74, 138.20, 137.99, 135.63, 134.63, 134.06, 133.23, 131.20, 129.29, 128.10, 127.88, 126.97, 125.65, 123.80, 123.67, 121.70, 120.08, 117.46, 115.50, 22.17, 18.29; ESI-MS *m/z* 404.4 (M + 1).

6,8-Dimethyl-*N*-(naphthalen-2-yl)-2-(pyridin-2-yl)quinoline-4-carboxamide (18). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine and 5,7-dimethylisatin was used instead of isatin. The crude compound was purified by chromatography using a mixture of hexanes and ethyl acetate (1:2). The overall yield was 25.1%; mp 226–227 °C; ¹H NMR (CDCl₃) δ 8.74–8.70 (m, 3H), 8.46 (d, *J* = 2.1 Hz, 1H), 8.16 (s, 1H), 8.00 (s, 1H), 7.92–7.82 (m, 4H), 7.63 (dd, *J* = 2.4, 8.7 Hz, 1H), 7.48 (m, 3H), 7.39–7.35 (m, 1H), 2.89 (s, 3H), 2.50 (s, 3H); ¹³C NMR (CDCl₃) δ 166.28, 155.94, 152.95, 148.91, 146.08, 141.72, 137.97, 137.58, 137.02, 135.18, 133.81, 132.69, 130.95, 128.97, 127.86, 127.62, 126.63, 125.32, 124.37, 124.14, 121.91, 121.65, 119.96, 117.25, 115.57, 21.95, 18.13; ESI-MS *m/z* 404.3 (M + 1).

6-Chloro-*N*-(naphthalen-2-yl)-2-(pyridin-4-yl)quinoline-4-carboxamide (19). The reaction conditions were the same as above except isatin was substituted with 5-chloroisatin. The overall yield was 8.3%; mp 225–226 °C; ¹H NMR (DMSO) δ 11.10 (s, 1H), 8.81 (d, *J* = 5.1 Hz, 2H), 8.64 (s, 1H), 8.56 (s, 1H), 8.34–8.31 (m, 3H), 8.26 (d, *J* = 9 Hz, 1H), 7.97–7.88 (m, 4H), 7.77 (d, *J* = 8.7 Hz, 1H), 7.54–7.45 (m, 2H); ¹³C NMR (DMSO) δ 164.6, 154.0, 150.5, 146.4, 144.6, 142.0, 136.1, 133.2, 132.8, 132.0, 131.2, 130.2, 128.4, 127.5, 126.5, 125.1, 124.6, 123.9, 121.3, 120.4, 118.2, 116.6; ESI-MS *m/z* 410.3 (M + 1), 382.4 (M – 28), 267.0 (M – 143), 239.2 (M – 171).

6-Chloro-*N*-(naphthalen-2-yl)-2-(pyridin-3-yl)quinoline-4-carboxamide (20). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and

5-chloroisatin was used instead of isatin. The overall yield was 3.2%; mp 228–229 °C; ¹H NMR (DMSO) δ 11.07 (s, 1H), 9.54 (s, 1H), 8.73 (d, *J* = 7.5 Hz, 2H), 8.61 (s, 1H), 8.56 (s, 1H), 8.30 (d, *J* = 2.7 Hz, 1H), 8.23 (d, *J* = 9 Hz, 1H), 7.97–7.88 (m, 4H), 7.78 (d, *J* = 8.4 Hz, 1H), 7.64–7.60 (m, 1H), 7.54–7.43 (m, 2H); ¹³C NMR (DMSO) δ 164.8, 154.5, 150.9, 148.6, 146.6, 141.9, 136.2, 134.9, 133.3, 132.4, 131.9, 131.2, 130.3, 128.6, 127.6, 126.7, 125.2, 124.2, 124.0, 120.6, 118.4, 116.7; ESI-MS *m/z* 410.2 (M + 1), 382.4 (M – 28), 267.0 (M – 143), 239.2 (M – 171).

6-Bromo-*N*-(naphthalen-2-yl)-2-(pyridin-4-yl)quinoline-4-carboxamide (21). The reaction conditions were the same as above except isatin was substituted with 5-bromoisoat. The overall yield was 5.6%; mp 208–209 °C; ¹H NMR (DMSO) δ 11.10 (s, 1H), 8.81 (d, *J* = 3.9 Hz, 2H), 8.63 (s, 1H), 8.56 (s, 1H), 8.47 (d, *J* = 2.1 Hz, 1H), 8.33 (d, *J* = 5.7 Hz, 2H), 8.18 (d, *J* = 8.7 Hz, 1H), 8.04 (dd, *J* = 2.1, 9.3 Hz, 1H), 7.92 (m, 3H), 7.77 (dd, *J* = 2.1, 8.7 Hz, 1H), 7.54–7.43 (m, 2H); ¹³C NMR (DMSO) δ 164.7, 154.1, 150.3, 146.6, 144.9, 141.9, 136.2, 133.9, 133.3, 132.1, 130.3, 128.5, 127.6, 127.5, 127.3, 126.6, 125.2, 125.1, 121.7, 121.5, 120.5, 118.4, 116.6; ESI-MS *m/z* 454.2 (M + 1), 426.3 (M – 28), 311.0 (M – 143), 283.2 (M – 171).

6-Bromo-*N*-(naphthalen-2-yl)-2-(pyridin-3-yl)quinoline-4-carboxamide (22). The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and 5-bromoisoat was used instead of isatin. The overall yield was 6.1%; mp 240–241 °C; ¹H NMR (DMSO) δ 11.07 (s, 1H), 9.54 (s, 1H), 8.74–8.71 (m, 2H), 8.61 (s, 1H), 8.56 (s, 1H), 8.45 (d, *J* = 2.1 Hz, 1H), 8.15 (d, *J* = 9 Hz, 1H), 8.03–7.88 (m, 4H), 7.77 (dd, *J* = 1.5, 8.7 Hz, 1H), 7.64–7.60 (m, 1H), 7.54–7.43 (m, 2H); ¹³C NMR (DMSO) δ 164.7, 154.5, 150.9, 148.6, 146.7, 141.7, 136.2, 134.8, 133.7, 133.3, 131.9, 130.3, 128.5, 127.6, 127.2, 126.6, 125.2, 124.6, 124.0, 121.1, 120.5, 118.3, 116.7; ESI-MS *m/z* 454.2 (M + 1), 426.3 (M – 28), 311.0 (M – 143), 283.1 (M – 171).

***N*-(Naphthalen-1-yl)-2-(pyridin-3-yl)quinoline-4-carboxamide (23).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 3-acetylpyridine and 1-naphthylamine was used instead of 2-naphthylamine to form the amide. The overall yield was 35.8%; mp 201–202 °C; ¹H NMR (CDCl₃) δ 9.27 (s, 1H), 8.67–8.61 (m, 2H), 8.47 (d, *J* = 7.8 Hz, 1H), 8.36 (d, *J* = 8.1 Hz, 1H), 8.21 (t, *J* = 7.8 Hz, 2H), 8.11 (s, 1H), 7.95–7.91 (m, 2H), 7.85–7.80 (m, 2H), 7.67–7.52 (m, 4H), 7.42–7.38 (m, 1H); ¹³C NMR (CDCl₃) δ 166.05, 154.16, 150.44, 148.93, 148.57, 143.31, 134.86, 134.27, 134.21, 131.71, 130.76, 130.35, 128.98, 128.19, 127.11, 126.75, 126.30, 125.77, 124.98, 123.74, 123.59, 121.33, 120.53, 116.19; ESI-MS *m/z* 376.5 (M + 1).

***N*-(Naphthalen-1-yl)-2-(pyridin-2-yl)quinoline-4-carboxamide (24).** The reaction conditions were the same as above except 4-acetylpyridine was substituted with 2-acetylpyridine and 1-naphthylamine was used instead of 2-naphthylamine to form the amide. The overall yield was 3.8%; mp 207–208 °C; ¹H NMR (DMSO) δ 10.99 (s, 1H), 8.87–8.81 (m, 2H), 8.68 (d, *J* = 7.8 Hz, 1H), 8.31–8.23 (m, 2H), 8.13–7.99 (m, 3H), 7.92–7.86 (m, 3H), 7.75 (t, *J* = 7.2 Hz, 1H), 7.64–7.57 (m, 4H); ¹³C NMR (DMSO) δ 167.15, 155.84, 155.32, 150.18, 148.39, 143.90, 138.35, 134.51, 133.63, 131.20, 130.51, 129.20, 128.93, 128.80, 127.21, 126.97, 126.92, 126.35, 126.02, 125.79, 125.04, 124.11, 123.53, 121.92, 117.14; ESI-MS *m/z* 376.5 (M + 1).

Difference Spectroscopy of Binding and Substrate Titrations. Difference spectra were collected using a double-beam Olis upgraded Amino DW-2000 spectrophotometer (Olis, Inc., Bogart, GA). The wavelength range encompassed 350–500 nm. Both sample and reference cuvettes contained purified 2C9 in 100 mM KPi buffer with 20% glycerol, pH 7.4.¹⁰ To obtain difference spectra, a baseline was recorded with 2C9 enzyme and buffer in both the sample and reference cuvettes followed by substrate titration into the sample cuvette, and the same volume of solvent was added to the reference cuvette. The difference spectra of samples containing purified 2C9 0.49 μM and substrates were scanned at 30 °C. Substrate concentrations were between 0.107 and 109.9 μM. Substrate binding curves were fit by nonlinear regression

with GraphPad Prism4 using equation $\Delta A = (B_{\max}[S])/(K_s + [S])$, where A is the absorbance difference, B_{\max} is the maximum absorbance difference, $[S]$ is the substrate concentration. The types of substrate-induced binding spectra with P450 heme were determined by the positions of wavelengths for peaks (λ_{\max}) and minima (λ_{\min}) on the spectra.²⁹

EPR Studies on Binding Modes. EPR spectra were recorded on a Bruker ESP-300e X-band spectrometer fitted with a liquid helium flow cryostat (ESR-9, Oxford Instruments). The ferric 2C9 samples were prepared in KPi buffer (100 mM, pH 7.4, 20% glycerol) and frozen in liquid nitrogen prior to spectral acquisition. The protein concentrations of samples were 18.6 μM (substrate-free form) and 18.5 μM (substrate-bound form), respectively, with a substrate concentration of 1 mM. Spectra were recorded at 20 K with microwave frequency of 9.614 GHz, microwave power of 10 mW, modulation amplitude of 16 G, modulation frequency of 100 kHz, receiver gain of 1×10^4 , conversion time of 168 ms, and time constant of 20 ms.

pKa Measurement. Data were collected on a diode array HP 845 UV-vis spectrophotometer. Samples were prepared by adding 500 μL of 0.1 mM substrate stock solution in DMSO to 500 μL of 0.1 M KPi buffer prepared at pH values between 1.04 and 10.03. As the pyridine nitrogen is protonated at lower pH, peaks emerged with λ_{\max} at 287 nm. Absorbance was plotted versus pH and fit to a sigmoidal function using GraphPad Prism4. Results agreed with MarvinSketch calculated values.

Automated Docking. Substrates were docked into the crystal structure of 2C9 1r9o³⁰ using MOE 2006.08 (Chemical Computing Group Inc. Montreal, Quebec, Canada). Compounds were sketched and underwent energy minimization as entries. Selected substrates were then fit into the active site cavity using the automatic docking program, exploring the conformational space associated with each rotatable bond. The parameters were as follows: receptor, receptor atoms; site, selected atoms (glycerol + flurbiprofen); ligand, sketched substrates; placement, alpha triangle; scoring, London dG; retain, 100. The parameters for inclusion of the residues by ligand interactions calculations in MOE were fields of H-bond, ionic, and solvent with a minimum score 10% prospectively. Only the substrate and amino acid residues within 4.5 Å were allowed to move during the minimization. The protein was frozen when the autodocking was performed. The water molecules were modeled within the crystal structure before the docking process was initiated. The force field for both docking and energy minimization were Hamiltonian, MMFF94x in MOE.

DFT Calculations. Density functional calculations were performed using Gaussian 03.³¹ The TPSS functional³² was used with the cc-pVDZ basis set with the effective core potential on iron,³³ the aug-cc-pVDZ on sulfur³⁴ and nitrogen³⁵ with diffuse d functions removed, and the cc-VDZ basis set on carbon and hydrogen.³⁶ The basis set and optimized geometries are available as Supporting Information. Solvation energies were calculated for the gas phase optimized geometries using the polarized continuum model self-consistent reaction field with the solvent set to water's dielectric constant. The heme model was the abbreviated heme with an S–H fifth ligand used by Shaik and co-workers.³⁷

Results

All quinoline-4-carboxamide analogues were synthesized by the route shown in Scheme 1. Results of the inhibition assays are shown in Table 1. From the experimental results, we saw a trend in binding affinity related to the positions of the type II accessible nitrogen in the pyridine moiety (Figure 1). Compounds with a pyridine nitrogen in the para position (relative to the linkage) generally exhibit higher binding affinity than those in the meta and ortho positions. In addition, compounds with an aromatic ring system at the carboxyamide moiety bind more tightly than those that lack a ring system (Figure 1, R = aromatic).

UV difference spectra were used to determine substrate binding modes and spectral dissociation constants K_s for the

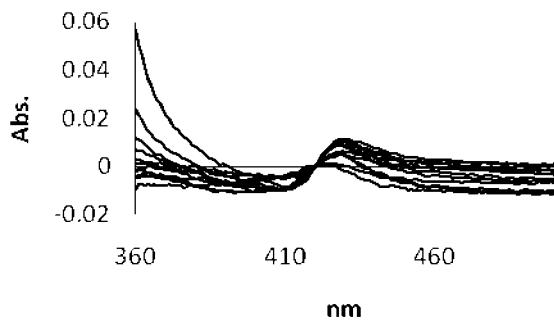


Figure 2. Binding spectrum and titration curves of compound **13**, $K_i = 0.325 \mu\text{M}$.

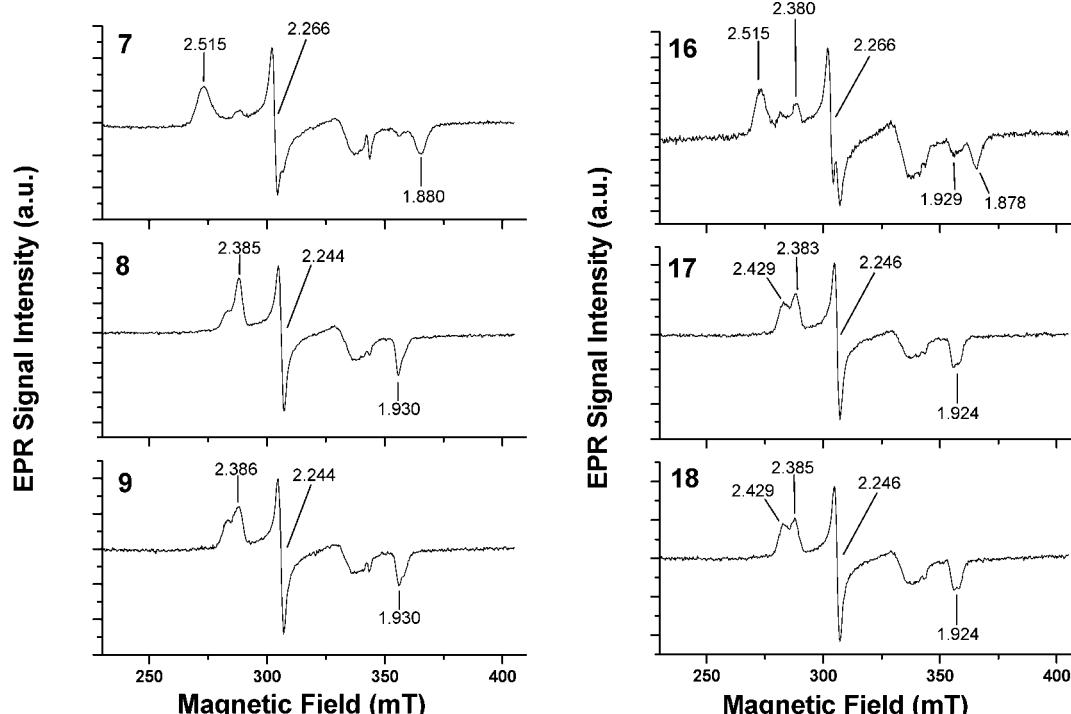
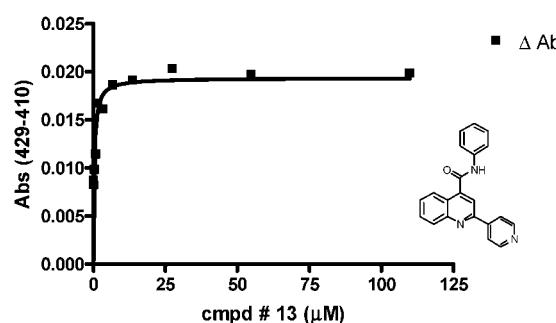


Figure 3. EPR spectra of 2C9 in the presence of substrates **7** (para), **8** (meta), **9** (ortho), **16** (para), **17** (meta), and **18** (ortho).

quinoline-4-carboxamide analogues shown in Table 1 whose absorbance did not interfere with the UV absorption of the titration. Of those analogues, **10** and **13** gave clear type II binding spectra (representative titration results are shown in Figure 2). Since the K_i values reported in Table 1 were obtained using baculosomes, we used the reconstituted 2C9 system to determine if purified enzyme gave different K_i values relative to the baculosomes preparations. The results gave K_i values for compound **10** of $113 \pm 6.85 \text{ nM}$ in the purified system and 125 nM in the baculosomes. For compound **13**, the K_i was $79.8 \pm 5.91 \text{ nM}$ in the purified system and 100 nM in the baculosome system. The results indicate that for these substrates the purified reconstituted system and the baculosomes system gave similar results. Compound **14** showed a peak at $\sim 415 \text{ nm}$ but no obvious trough in the difference spectra, which implies that **14** exhibits mixed type II and type I binding modes.

Since EPR measurements are not hindered by the substrate absorbing in the Soret band range, additional compounds could be examined for the binding mode. Compounds **7**, **8**, **9**, **16**, **17**, **18**, **19**, and **21** were studied by EPR to probe binding modes (Figure 3 and 4). From these results, analogues with the pyridine nitrogen in the para position to the linkage (**7**, **16**, **19**, and **21**), EPR spectra showed a low-spin shift which was absent with compounds having nitrogen in the meta or ortho positions. The

EPR spectrum of substrate-free 2C9 had g -values at 2.42/2.38, 2.24, and 1.92 (data not shown), and the type II binding substrate bound 2C9 spectrum had g -values at 2.51, 2.26, and 1.88. This result suggests that the para nitrogen is coordinated to the heme iron, while the meta and ortho compounds are not.

To determine which portion of the total binding energy depends on pure type II interaction (as opposed to apoprotein interactions), the density functional method TPSS was used for the core structure of the three pyridinylquinolines (ortho, meta, and para pyridines, Figure 5), and the differences in energies for nitrogen substitution ortho, meta, and para from the linkage to quinoline were determined. The results indicate that the ortho nitrogen linkage has a much longer N–Fe bond and significantly distorts the heme from planarity. In fact, this structure was so distorted that the geometry could not be minimized. The meta and para linkage both have 2.1 \AA N–Fe bond lengths with only a slight distortion in the planarity of the heme. Electronically, the para linkage has the strongest interaction energy by 0.6 kcal/mol ; however, this is not a significant difference given the level of computation.

To determine if differences in solvation energy might influence the overall binding energy, a series of ortho, meta, and para linkages were used to calculate the solvation energies for each core structure using a polarized continuum self-

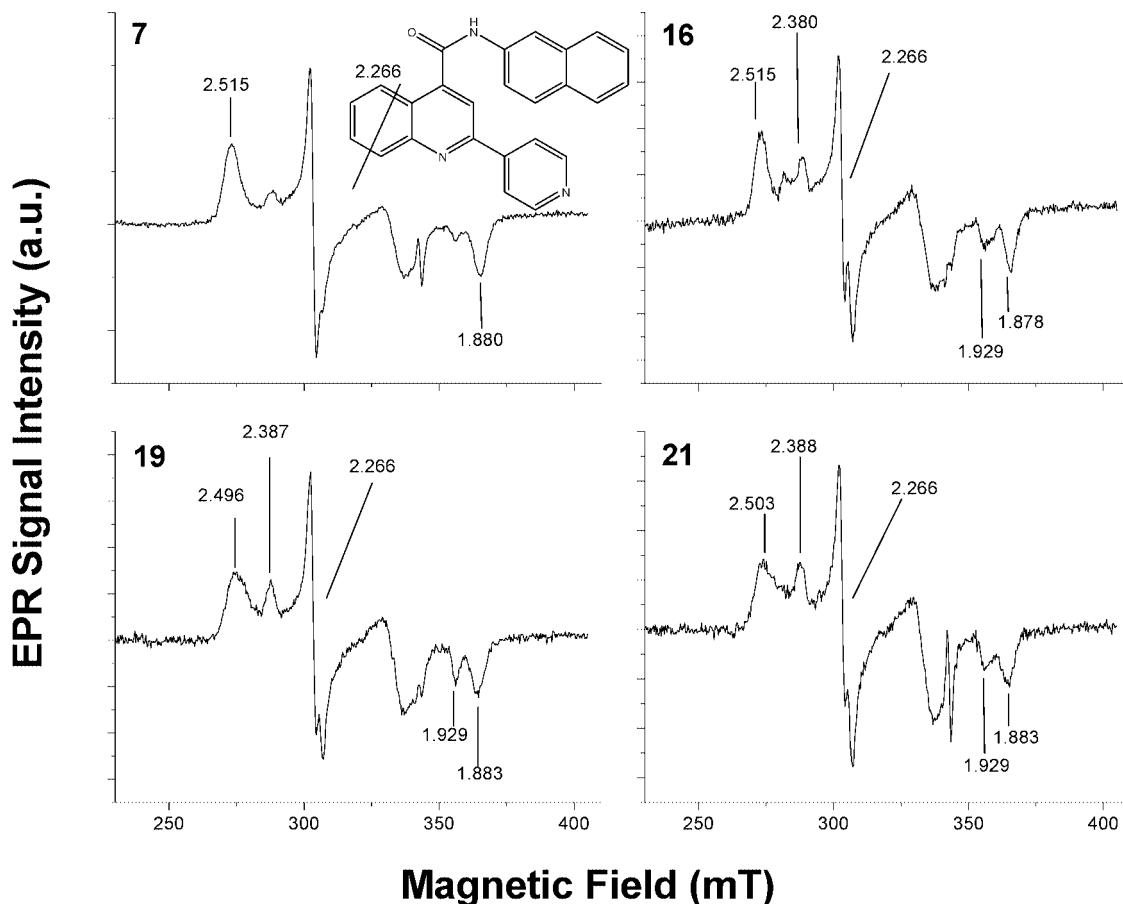


Figure 4. EPR spectra of 2C9 in the presence of substrates **7** (para), **16** (para), **19** (para), and **21** (para).

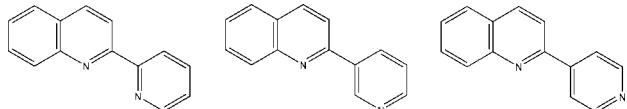


Figure 5. Core structures used to determine nitrogen–iron coordination energies and solvation energies.

consistent reaction field model as implemented in Gaussian 03. Results indicate insignificant differences in solvation energy between the meta and para isomer, while the ortho isomer was around 2 kcal/mol less stabilized by solvent. This difference in solvent interaction is also reflected in the measured differences in pK_a values for the ortho compound **18** of 3.05 versus nearly identical values of 4.27 and 4.30 for para (**16**) and meta (**17**), respectively. These results indicate that the difference in binding affinity for the compounds with the ortho linkage may be complicated by differences in solvation.

Discussion

Understanding the mechanism of type I and type II substrate binding modes in P450 drug metabolizing isoforms is critical to optimizing drug design and reducing drug toxicity. Metabolic stability, enzyme inhibition, metabolite regioselectivity, and formation of reactive metabolites are all functions of binding modes. It has been shown that substrates which are capable of type II binding are more potent inhibitors than those capable of only type I binding.³⁸ In addition, potent inhibition is possible when coordination between the heteroatom (nitrogen atom) lone pair electrons and the ferric heme iron is combined with apoprotein interactions (lipophilic and electrostatic).¹¹ Type II binding slows the rate of P450-mediated oxidation, increasing

substrate metabolic stability as well as the potency of inhibition. Therefore, elucidating how type II interactions can be used to optimize drugs metabolic properties without increasing drug toxicity is necessary to optimize drug design.

In this study we investigated the potential increase in binding affinity that can be attributed to type II binding in the absence of other binding features. To this end, we synthesized a series of quinoline-4-carboxamide analogues that differ in the placement of nitrogen capable of forming a type II complex with the heme iron of 2C9 (Table 1). Each set of three compounds differed only in the position of their linkage and are assumed to have almost identical solvation energies allowing for the determination of the nitrogen coordination binding. One caveat is that solvation energy calculations and pK_a measurements show that interpretation of changes in binding affinity for the ortho compounds may be complicated by differences in solvation relative to the meta and para linkages.

The effects of varying the position of the nitrogen atom in the pyridine ring on the binding affinity of quinoline-4-carboxamide analogues to 2C9 are shown in Table 1 and Figure 6. In general, compounds with a para nitrogen in the pyridine ring bind more tightly than meta or ortho compounds; compounds with meta nitrogens are better binders than ortho compounds. Para compounds generally bind to 2C9 with K_i between 9 and 125 nM except for compounds **1** and **4**, which have K_i values of 9.94 and 3.67 μ M, respectively. The binding affinity of para compounds are increased by a factor of up to 4200-fold compared with meta compounds (such as **21** and **22**) and a factor of up to 1900-fold compared with ortho compounds (such as **16** and **18**). Meta compounds overall have a larger range of binding affinities to 2C9, having K_i values from 0.15

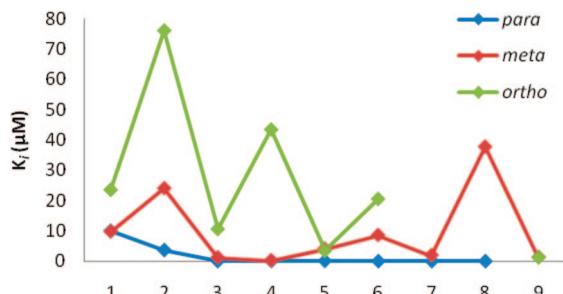


Figure 6. K_i versus nitrogen atom in the para, meta, and ortho positions toward the linkage of nine different groups of compounds indicating that, in general, the order of tightness of binding is para > meta > ortho (Table 1).

Table 2. B3LYP/cc-pVDZ Substrate Energies and Heme–Substrate Complexes for Core Structures Shown in Figure 5

linkage	substrate energy ^a	type II energy ^b	solvation energy ^c
para	−649.207	−2135.127	−10.23
meta	−649.208	−2135.126	−9.87
ortho	−649.344	not stable	−8.03

^a Energies in Hartrees for the para, meta, and ortho linked core structures in Figure 5. ^b Energies in Hartrees for each core structure coordinated to the heme. ^c PCM solvation energies in kcal/mol for the different linkages with the dielectric set to that of water.

Table 3. List of UV and EPR Spectral Results for Selected Compounds

compound	K_i (μM)	binding type	
		UV min (nm)/max (nm)	EPR
1	9.94	type II 410/429	type II
2	9.74	mixed -/413	type I
7	0.033	ND ^b	type II
8	1.15	ND ^a	type I
9	10.67	ND ^a	type I
10	0.125	type II 413/428	ND ^a
13	0.10	type II 410/431	ND ^a
14	4.06	mixed -/417	ND ^a
16	0.011	ND ^c	type II/type I
17	8.55	ND ^c	type I
18	>20.6	ND ^c	type I
19	0.012	ND ^b	type II/type I
21	0.009	ND ^b	type II/type I

^a Data not collected. ^b Substrate absorbance prevented characterization. ^c Spectrum is not clear.

to $37.85 \mu\text{M}$, and these compounds can be 300-fold tighter binders than ortho compounds. In general, ortho compounds bind less tightly than those of their para and meta counterparts. K_i values of ortho compounds ranged from 76.1 – $1.32 \mu\text{M}$. The most likely explanation for these results is that the para compounds are capable of stronger type II binding interaction than the meta compounds, which are capable of stronger type II interactions than the ortho compounds.

To determine if type II binding is responsible for the differences in binding affinity, we used both UV and EPR spectroscopy (Table 3) to determine substrate binding modes of several compounds from Table 1. Binding mode analysis using UV spectroscopy was limited to only compounds that did not absorb in the region of interest, 350–500 nm. UV difference spectra results for para compounds (**1**, **10**, and **13**) show typical type II binding spectra, with peaks at ~ 430 nm and a trough at ~ 410 nm. The meta compounds (**2** and **14**), however, show what appears to be mixed type I and type II binding in the UV difference spectra where peaks are at ~ 418 nm but not an obvious trough. The ortho compound (**15**) does not give either a type I or type II binding spectra.

EPR studies on para compounds **7**, **16**, **19**, and **21** suggest that the para nitrogen atom on the pyridine moiety is coordinated to the P450 iron. The low-spin *g*-value shifts of 1.91 to 1.88 and 2.45 to 2.51 observed for these compounds are typical of the low-spin shift commonly seen in type II binding substrates (normal nitrogen donors) in other P450 enzymes.^{8,39} However, compounds **16**, **19**, and **21** also show a low spin signal at 2.38 that is characteristic of a nonsubstrate bound EPR spectrum (Figure 4). The coexistence of both the shifts and the unbound signal at 2.38 for compounds **16**, **19**, and **21** suggests compound **7** is able to bind pure type II while the others bind in the mixed mode. The difference in binding modes of these para compounds may be attributed to the increase in steric bulk (and hydrophobicity) of compounds **16**, **19**, and **21** relative to compound **7**. While all are tight binders, increasing steric bulk may interfere with formation of a nitrogen–ferric complex allowing for the mixed binding mode seen in the EPR. Overall, UV and EPR studies support the hypothesis that the pyridine rings with the para-substituted nitrogen have strong type II binding interactions. The meta compounds appear to be able to coordinate to the heme to a lesser extent, while the ortho compounds do not appear to coordinate to the heme.

Interestingly, the UV difference spectra appear to provide a more sensitive test of substrate binding modes than EPR spectra. UV difference spectrum gathered on meta compound **2** showed what may be interpreted as mixed type I and type II binding that was not apparent in the EPR spectrum. The disagreement between the UV and EPR spectra of meta compounds could be a sensitivity issue or other artifacts requiring additional investigation.

The spectral dissociation constants (K_s) for compounds **10** and **13** (1.28 and $0.325 \mu\text{M}$) were higher than the inhibitor dissociation constant, K_i , (0.125 and $0.1 \mu\text{M}$) determined by IC₅₀ experiments. These discrepancies are not unusual^{40,41} and may be attributed to the different methods used to measure each value. K_s was measured as the type II binding substrate dissociation constant when the heme iron was in the +3 oxidation state and the amount of spin state change between high and low was dependent on the equilibrium of the nature of the nonsubstrate bound protein. On the other hand, K_i was determined as the overall inhibitor dissociation constant where the catalytic cycle was completed and the iron is in equilibrium between the +3 and +2 oxidation states, allowing the inhibitor to bind in either type I or type II modes. Mixed substrate binding and heme iron oxidation state equilibria further complicate the elucidation of the substrate binding modes within the active site. Another possible reason that the K_s was higher than K_i was that the substrate heme coordination was incomplete. Incomplete heme coordination has been documented by Locuson et al. 2007.¹⁰

One potential complication of this interpretation would arise if the para, meta, and ortho compounds had different pK_a values. To determine if the nitrogen atom in the pyridine ring was differentially protonated for the ortho, meta and para substituted pyridines in the physiological pH, the pK_a values of all analogues were predicted using MarvinSketch. To ensure that these predictions were reliable, pK_a values were experimentally determined for compounds **16**, **17**, and **18**. The results from computational prediction/experimental determination were 4.15/4.27, 3.44/4.30, and 3.36/3.05 for para, meta, and ortho, respectively. All the compounds have a predicted pK_a less than 4.3 for the pyridine nitrogen, which indicates that at physiological pH, the analogues are free to coordinate with the heme iron. In addition, a lower pK_a corresponds to greater delocalization of the aromatic ring.

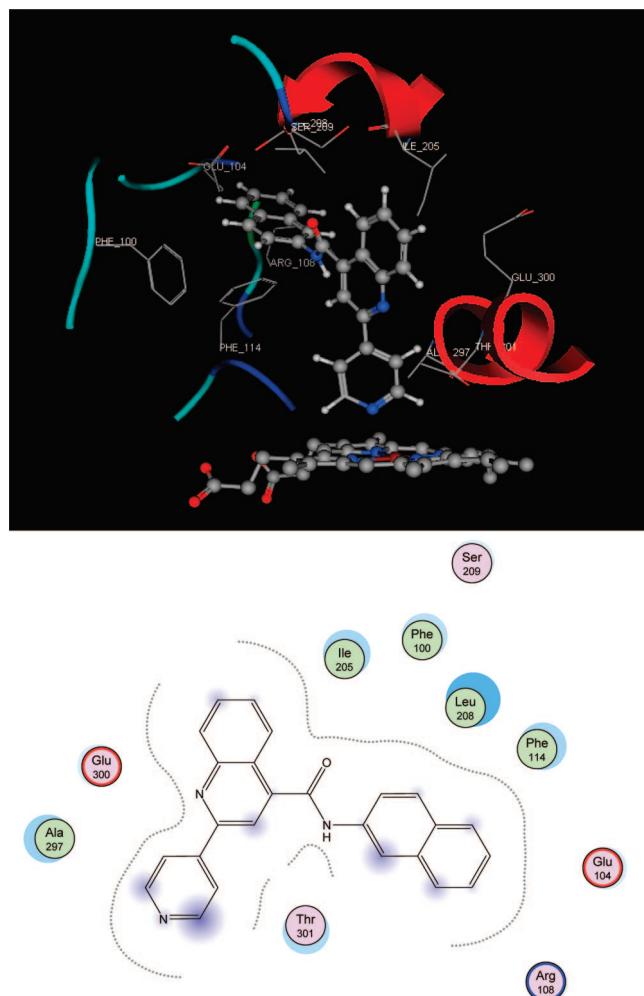


Figure 7. Proposed type II binding mode with compound **7** using the flurbiprofen complex of 2C9dH (Protein Data Bank code 1r9o), on the top. A 2D diagram of ligand–residue interactions is presented on the bottom.

ization of the lone pair in the aromatic system. Delocalization decreases the ability of an sp^2 nitrogen to type II binding. The piperidine nitrogen on compounds **4**, **5**, and **6**, which are predicted to have pK_a values of ~ 9.1 , and since these values are well above physiological pH, they would exist in their protonated forms. In general, positively charged compounds do not bind strongly to 2C9, which may explain the overall low affinity of compounds **4**, **5**, and **6** for the enzyme.

To determine if differential interactions of the para, meta, and ortho compounds result from different heme interactions, quantum calculations of the binding energies of the different compounds were determined (Table 2). The results indicate that the ortho substituted pyridines do not strongly interact with the heme. However, the para and meta compounds interact with essentially the same energy. These results suggest the meta compounds may have less favorable apoprotein interactions than the para substituted compounds. Further, these computational results suggest that active site residues play an important role in determining the affinity for the meta and para compounds' ability to form type II complexes. For meta and ortho compounds, interactions with the apoprotein rather than just the electrostatic interaction with the heme appear to have the most significant influence on binding affinity versus para compounds.

Crystal structures that show type II binding interactions include P450cam with nicotine, CYP51 with 4-phenylimidazole

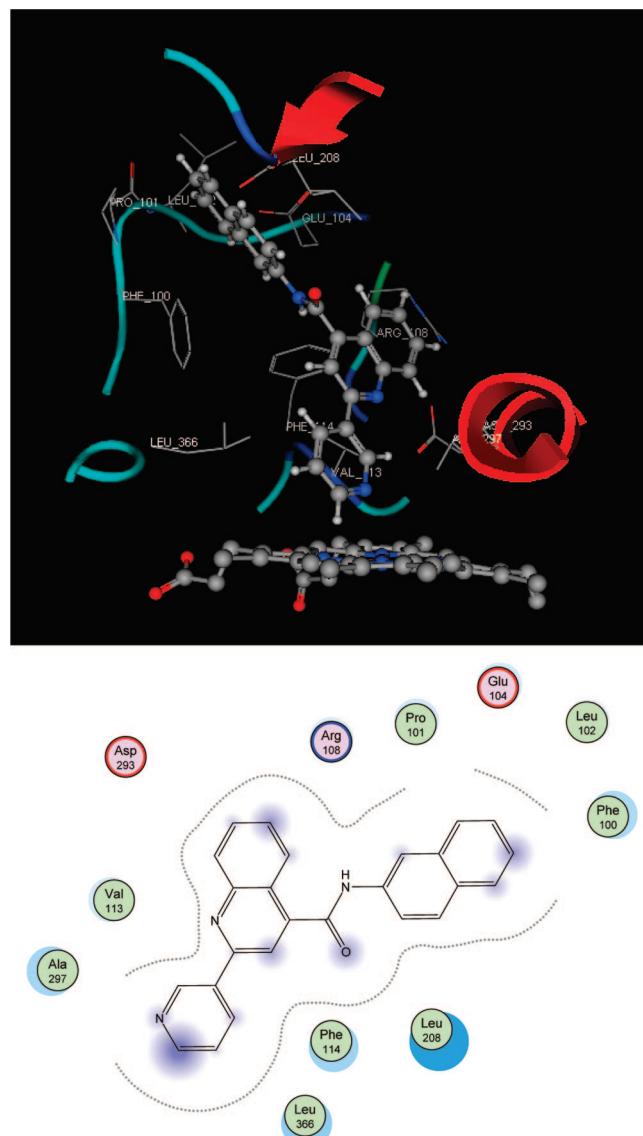


Figure 8. Proposed implausible type II binding mode with compound **8** using the flurbiprofen complex of 2C9dH (Protein Data Bank code 1r9o).

(4-PI) and fluconazole (FLU), P450eryF with ketoconazole, and CYP2B4 with bifonazole.^{42–45} Generally, these structures show the heterocyclic substrates type II bound to the heme iron with the remainder of the molecule bent away from the I-helix in the active site pocket and the essential conformational changes of apoprotein are necessary for substrates binding. With these substrate–enzyme binding interactions in mind, we applied computational modeling methods to further visualize the overall substrate binding interactions.

In an attempt to ascertain specific substrate–protein interactions, we used automated docking on two of our tight binders, the quinoline-4-carboxamide analogues **7** and **8** (Figures 7 and 8). Both of these compounds contain naphthalene functionalized amides and differ only in the position of the sp^2 nitrogen in the pyridine moiety. Indeed, all compounds from Table 1 that contain aromatic amides (especially naphthalene) bind more tightly than substrates with nonaromatic amides. This suggests that in addition to type II coordination, the aromatic amide systems may interact favorably with active site residues. Docking of type II binder **7** into the crystal structure of 2C9 is relatively straightforward since the nitrogen–heme coordination

is known. This important feature of the nitrogen position on pyridine is one of the major characteristics on substrate structures that determine overall binding. For compound **7**, a direct coordination between the heme and the para pyridine nitrogen is shown and this pyridine ring system is tilted away from the I-helix. In this orientation, the naphthalene ring faces Phe 114, allowing an edge-to-face π interaction. In addition, the positively charged side chain of Arg 108 is oriented in such a way to allow interaction with the naphthalene ring through parallel stacking geometry. Presumably, a π -cation interaction from the positively charged Arg 108 side chain with the aromatic ring of amide allows for favorable enthalpic contributions of van der Waals and electrostatic interactions. This type of energetically favorable π -cation interaction has been described in various biological systems and recognition processes.^{46,47} In addition, the quinoline ring portion of **7** is located in a hydrophobic pocket surrounded by Phe 476, Leu 208, Ile 205, and Leu 362. This conformation allows for potential π - π interactions between the quinoline ring and Phe 476.

The proposed binding orientation for compound **8** does not have the nitrogen atom coordinated to the heme iron. This may be a result of steric interference with the I-helix, even though all the other ligand-residue interactions are similar to those for compound **7**. These docking results suggest the primary difference in binding affinity between the meta and para compounds is a result of steric interaction with the I-helix resulting in diminished or nonexistent type II binding.

Conclusion

Type II interactions involving coordination of a pyridine nitrogen to the heme results in an up to 4200-fold increase in binding affinity. These significant increases in affinity can be modulated by changing the position of the nitrogen relative to the substituents and could be used to dial-in affinities for P450 enzymes to minimize drug-drug interactions, while maintaining very similar reactivity and solubility in a potential drug.

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Supporting Information Available: The cc-aug-pVDZ basis set and effective core potential used for calculations and tables of Cartesian coordinates for geometries used for Table 2 and the chromatographic data for target compounds. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- Locuson, C. W.; Wahlstrom, J. L.; Rock, D. A.; Rock, D. A.; Jones, J. P. A new class of CYP2C9 inhibitors: Probing 2C9 specificity with high-affinity benzboromarone derivatives. *Drug Metab. Dispos.* **2003**, *31*, 967–971.
- Locuson, C. W.; Suzuki, H.; Rettie, A. E.; Jones, J. P. Charge and substituent effects on affinity and metabolism of benzboromarone-based CYP2C19 inhibitors. *J. Med. Chem.* **2004**, *47*, 6768–6776.
- Locuson, C. W.; Rock, D. A.; Jones, J. P. Quantitative binding models for CYP2C9 based on benzboromarone analogues. *Biochemistry* **2004**, *43*, 6948–6958.
- Rettie, A. E.; Jones, J. P. Clinical and toxicological relevance of CYP2C9: Drug-drug interactions and pharmacogenetics. *Annu. Rev. Pharmacol. Toxicol.* **2005**, *45*, 477–494.
- Hudelson, M. G.; Jones, J. P. Line-walking method for predicting the inhibition of P450 drug metabolism. *J. Med. Chem.* **2006**, *49*, 4367–4373.
- Hudelson, M. G.; Ketkar, N. S.; Holder, L. B.; Carlson, T. J.; Peng, C. C.; Waldher, B. J.; Jones, J. P. High confidence predictions of drug-drug interactions: predicting affinities for cytochrome P450 2C9 with multiple computational methods. *J. Med. Chem.* **2008**, *51*, 648–654.
- Schenkman, J. B. Studies on the nature of the type I and type II spectral changes in liver microsomes. *Biochemistry* **1970**, *9*, 2081–2091.
- Kumaki, K.; Sato, M.; Kon, H.; Nebert, D. W. Correlation of type I, type II, and reverse type I difference spectra with absolute changes in spin state of hepatic microsomal cytochrome P-450 iron from five mammalian species. *J. Biol. Chem.* **1978**, *253*, 1048–1058.
- Schenkman, J. B.; Remmer, H.; Estabrook, R. W. Spectral studies of drug interaction with hepatic microsomal cytochrome. *Mol. Pharmacol.* **1967**, *3*, 113–123.
- Locuson, C. W.; Hutzler, J. M.; Tracy, T. S. Visible spectra of type II cytochrome P450-drug complexes: evidence that “incomplete” heme coordination is common. *Drug Metab. Dispos.* **2007**, *35*, 614–622.
- Chiba, M.; Jin, L.; Neway, W.; Vacca, J. P.; Tata, J. R.; Chapman, K.; Lin, J. H. P450 interaction with HIV protease inhibitors: relationship between metabolic stability, inhibitory potency, and P450 binding spectra. *Drug Metab. Dispos.* **2001**, *29*, 1–3.
- Roberts, A. G.; Campbell, A. P.; Atkins, W. M. The thermodynamic landscape of testosterone binding to cytochrome P 450 3A4: Ligand binding and spin state equilibria. *Biochemistry* **2005**, *44*, 1353–1366.
- LeLean, J. E.; Moon, N.; Dunham, W. R.; Coon, M. J. EPR spectrometry of cytochrome P450 2B4: Effects of mutations and substrate binding. *Biochem. Biophys. Res. Commun.* **2000**, *276*, 762–766.
- Tsai, R.; Yu, C. A.; Gunsalus, I. C.; Peisach, J.; Blumberg, W.; Orme-Johnson, W. H.; Beinert, H. Spin-state changes in cytochrome P-450cam on binding of specific substrates. *Proc. Natl. Acad. Sci. U.S.A.* **1970**, *66*, 1157–1163.
- Chiba, M.; Tang, C.; Neway, W. E.; Williams, T. M.; Desolms, S. J.; Dinsmore, C. J.; Wai, J. S.; Lin, J. H. P450 interaction with farnesyl-protein transferase inhibitors. Metabolic stability, inhibitory potency, and P450 binding spectra in human liver microsomes. *Biochem. Pharmacol.* **2001**, *62*, 773–776.
- Johnston, S. R.; Martin, L. A.; Head, J.; Smith, I.; Dowsett, M. Aromatase inhibitors: combinations with fulvestrant or signal transduction inhibitors as a strategy to overcome endocrine resistance. *J. Steroid Biochem. Mol. Biol.* **2005**, *95*, 173–181.
- Denton, T. T.; Zhang, X.; Cashman, J. R. Nicotine-related alkaloids and metabolites as inhibitors of human cytochrome P-450 2A6. *Biochem. Pharmacol.* **2004**, *67*, 751–756.
- Sellers, E. M.; Kaplan, H. L.; Tyndale, R. F. Inhibition of cytochrome P450 2A6 increases nicotine’s oral bioavailability and decreases smoking. *Clin. Pharmacol. Ther.* **2000**, *68*, 35–43.
- Ekins, S.; Mankowski, D. C.; Hoover, D. J.; Lawton, M. P.; Treadway, J. L.; Harwood, H. J., Jr. Three-dimensional quantitative structure-activity relationship analysis of human CYP51 inhibitors. *Drug Metab. Dispos.* **2007**, *35*, 493–500.
- Gibbons, G. F. The role of cytochrome P450 in the regulation of cholesterol biosynthesis. *Lipids* **2002**, *37*, 1163–1170.
- Zhang, W.; Ramamoorthy, Y.; Kilicarslan, T.; Nolte, H.; Tyndale, R. F.; Sellers, E. M. Inhibition of cytochrome P450 by antifungal imidazole derivatives. *Drug Metab. Dispos.* **2002**, *30*, 314–318.
- Zarn, J. A.; Bruschweiler, B. J.; Schlatter, J. R. Azole fungicides affect mammalian steroidogenesis by inhibiting sterol 14 alpha-demethylase and aromatase. *Environ. Health Perspect.* **2003**, *111*, 255–261.
- Evans, W. E.; Relling, M. V. Pharmacogenomics: translating functional genomics into rational therapeutics. *Science* **1999**, *286*, 487–491.
- Rettie, A. E.; Jones, J. P. Clinical and toxicological relevance of Cyp2c9: Drug-drug interactions and pharmacogenetics. *Annu. Rev. Pharmacol. Toxicol.* **2005**, *45*, 477–494.
- Rushmore, T. H.; Reider, P. J.; Slaughter, D.; Assang, C.; Shou, M. Bioreactor systems in drug metabolism: synthesis of cytochrome P450-generated metabolites. *Metab. Eng.* **2000**, *2*, 115–125.
- BD Biosciences. 4'-Hydroxydiclofenac standard. BD Biosciences: San Jose, CA, 2005.
- Shaw, P. M.; Hosea, N. A.; Thompson, D. V.; Lenius, J. M.; Guengerich, F. P. Reconstitution premixes for assays using purified recombinant human cytochrome P450, NADPH-cytochrome P450 reductase, and cytochrome b5. *Arch. Biochem. Biophys.* **1997**, *348*, 107–115.
- Locuson, C. W.; Gannett, P. M.; Tracy, T. S. Heteroactivator effects on the coupling and spin state equilibrium of CYP2C9. *Arch. Biochem. Biophys.* **2006**, *449*, 115–129.
- Jefcoate, C. R. Measurement of substrate and inhibitor binding to microsomal cytochrome P-450 by optical-difference spectroscopy. *Methods Enzymol.* **1978**, *52*.
- Wester, M. R.; Yano, J. K.; Schoch, G. A.; Yang, C.; Griffin, K. J.; Stout, C. D.; Johnson, E. F. The structure of human cytochrome P4502C9 complexed with flurbiprofen at 2.0-angstrom resolution. *J. Biol. Chem.* **2004**, *279*, 35630–35637.
- Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Montgomery, J. A., Jr.; Vreven, T.; Kudin, K. N.; Burant, J. C.; Millam, J. M.; Iyengar, S. S.; Tomasi, J.; Barone, V.; Mennucci, B.; Cossi, M.; Scalmani, G.; Rega, N.; Petersson, G. A.;

Nakatsuji, H.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Klene, M.; Li, X.; Knox, J. E.; Hratchian, H. P.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Ayala, P. Y.; Morokuma, K.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Zakrzewski, V. G.; Dapprich, S.; Daniels, A. D.; Strain, M. C.; Farkas, O.; Malick, D. K.; Rabuck, A. D.; Raghavachari, K.; Foresman, J. B.; Ortiz, J. V.; Cui, Q.; Baboul, A. G.; Clifford, S.; Cioslowski, J.; Stefanov, B. B.; Liu, G.; Liashenko, A.; Piskorz, P.; Komaromi, I.; Martin, R. L.; Fox, D. J.; Keith, T.; Al-Laham, M. A.; Peng, C. Y.; Nanayakkara, A.; Challacombe, M.; Gill, P. M. W.; Johnson, B.; Chen, W.; Wong, M. W.; Gonzalez, C.; Pople, J. A. *Gaussian 03*, revision C.02; Gaussian, Inc.: Wallingford, CT, 2004.

(32) Staroverov, V. N.; Scuseria, G. E.; Tao, J. M.; Perdew, J. P. Comparative assessment of a new nonempirical density functional: Molecules and hydrogen-bonded complexes. *J. Chem. Phys.* **2003**, *119*, 12129–12137.

(33) Peterson, K. A.; Dolg, M. Unpublished results.

(34) Woon, D. E.; Dunning, T. H. Gaussian-basis sets for use in correlated molecular calculations 0.3. The atoms aluminum through argon. *J. Chem. Phys.* **1993**, *98*, 1358–1371.

(35) Kendall, R. A.; Dunning, T. H.; Harrison, R. J. Electron-affinities of the 1st-row atoms revisited—systematic basis-sets and wave-functions. *J. Chem. Phys.* **1992**, *96*, 6796–6806.

(36) Dunning, T. H. Gaussian basis sets for use in correlated molecular calculations. I. The atoms boron through neon and hydrogen. *J. Chem. Phys.* **1989**, *90*, 1007–1023.

(37) Cho, K. B.; Moreau, Y.; Kumar, D.; Rock, D. A.; Jones, J. P.; Shaik, S. Formation of the active species of cytochrome P450 by using iodosylbenzene: A case for spin-selective reactivity. *Chem.—Eur. J.* **2007**, *13*, 4103–4115.

(38) Schenkman, J. B.; Sligar, S. G.; Cinti, D. L. Substrate interaction with cytochrome P-450. *Pharmacol. Ther.* **1981**, *12*, 43–71.

(39) Dawson, J. H.; Andersson, L. A.; Sono, M. Spectroscopic investigations of ferric cytochrome P-450-CAM ligand complexes. Identification of the ligand trans to cysteinate in the native enzyme. *J. Biol. Chem.* **1982**, *257*, 3606–3617.

(40) Verras, A.; Kuntz, I. D.; Ortiz de Montellano, P. R. Computer-assisted design of selective imidazole inhibitors for cytochrome p450 enzymes. *J. Med. Chem.* **2004**, *47*, 3572–3579.

(41) Ahlstrom, M. M.; Zamora, I. Characterization of type II ligands in CYP2C9 and CYP3A4. *J. Med. Chem.* **2008**, *51*, 1755–1763.

(42) Strickler, M.; Goldstein, B. M.; Maxfield, K.; Shireman, L.; Kim, G.; Matteson, D. S.; Jones, J. P. Crystallographic studies on the complex behavior of nicotine binding to P450cam (CYP101). *Biochemistry* **2003**, *42*, 11943–11950.

(43) Podust, L. M.; Poulos, T. L.; Waterman, M. R. Crystal structure of cytochrome P450 14alpha-sterol demethylase (CYP51) from *Mycobacterium tuberculosis* in complex with azole inhibitors. *Proc. Natl. Acad. Sci. U.S.A.* **2001**, *98*, 3068–3073.

(44) Cupp-Vickery, J. R.; Garcia, C.; Hofacre, A.; McGee-Estrada, K. Ketoconazole-induced conformational changes in the active site of cytochrome P450eryF. *J. Mol. Biol.* **2001**, *311*, 101–110.

(45) Zhao, Y.; White, M. A.; Muralidhara, B. K.; Sun, L.; Halpert, J. R.; Stout, C. D. Structure of microsomal cytochrome P450 2B4 complexed with the antifungal drug bifonazole: iInsight into P450 conformational plasticity and membrane interaction. *J. Biol. Chem.* **2006**, *281*, 5973–5981.

(46) Pellequer, J.-L.; Zhao, B.; Kao, H.-I.; Bell, C. W.; Li, K.; Li, Q. X.; Karu, A. E.; Roberts, V. A. Stabilization of bound polycyclic aromatic hydrocarbons by a p-cation interaction. *J. Mol. Biol.* **2000**, *302*, 691–699.

(47) Dougherty, D. A.; Stauffer, D. A. Acetylcholine binding by a synthetic receptor: implications for biological recognition. *Science (Washington, DC, U.S.)* **1990**, *250*, 1558–1560.

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